

Biochemical Pharmacology

Biochemical Pharmacology 64 (2002) 883-888

Nuclear factor-kappa B and cancer: its role in prevention and therapy

Alok C. Bharti, Bharat B. Aggarwal*

Cytokine Research Section, Department of Bioimmunotherapy, M. D. Anderson Cancer Center, University of Texas, Box 143, 1515 Holcomb Boulevard, Houston, TX 77030, USA

Received 14 February 2002; accepted 12 April 2002

Abstract

Cancer is a hyperproliferative disorder in which invasion and angiogenesis lead to tumor metastasis. Several genes that mediate tumorigenesis and metastasis are regulated by a nuclear transcription factor, nuclear factor kappa B (NF- κ B). A heterotrimeric complex consisting of p50, p65, and IkBa, NF- κ B is present in its inactive state in the cytoplasm. When NF- κ B is activated, IkBa is degraded and p50-p65 heterodimer is translocated to the nucleus, binds the DNA (at the promoter region), and activates gene. Research within the last few years has revealed that NF- κ B is activated by carcinogens, tumor promoters, inflammatory cytokines, and by chemotherapeutic agents. The activation of NF- κ B can suppress apoptosis, thus promoting chemoresistance and tumorigenesis. Interestingly, however, most chemopreventive agents appear to suppress the activation of the NF- κ B through inhibition of NF- κ B signaling pathway. These chemopreventive agents also sensitize the tumors to chemotherapeutic agents through abrogation of NF- κ B activation. Overall, these observations suggest that NF- κ B is an ideal target for chemoprevention and chemosensitization. This article reviews evidence supporting this hypothesis.

© 2002 Elsevier Science Inc. All rights reserved.

Keywords: Nuclear factor-κB; Chemoprevention; Chemosensitizaion

1. Introduction

From extensive research during last half of the century, the common saying that "you are what you eat" is becoming increasingly accepted. To prevent the onset of both cancer and cardiovascular diseases, the National Institute of Health in the United States has recommended a high-fiber, low-fat diet, consisting of more fruits and vegetables. The importance of the diet is further suggested by the epidemiological evidence that certain cancers (e.g.

breast, prostate, colon, and lung) are more prevalent in the developed countries than in the developing countries, most likely because of differences in dietary constituents. The molecular mechanism by which diet mediates its preventive and therapeutic effects is an active area of current research. We propose that there are constituents of the every-day diet that regulate the activity of certain transcription factors such as NF-κB that play a critical role in carcinogenesis.

NF-κB is a transcription factor discovered by Sen and Baltimore in 1986, in the kappa light chain of immunoglobulins in B cells [1]. Research over the last few years has revealed that NF-κB is an inducible and ubiquitously expressed transcription factor for genes involved in cell survival, cell adhesion, inflammation, differentiation, and growth [2–5]. Active NF-κB complexes are dimers of various combinations of the Rel family of polypeptides consisting of p50 (NF-κB1), p52 (NF-κB2), c-Rel, v-rel, Rel A (p65), and Rel B (see [6,7]). These proteins share a conserved 300 amino acid region within their amino termini, termed RHD, that is responsible for DNA binding, dimerization, nuclear translocation, and interaction with heterologous transcription factors. Although all Rel family

^{*}Corresponding author. Tel.: +1-713-792-3503; fax: 1+713-794-1613. *E-mail address:* aggarwal@mdanderson.org (B.B. Aggarwal).

Abbreviations: NF-κB, nuclear factor kappa B; RHD, Rel homology domain; IκB, inhibitor of NF-κB; IKK, IκBα kinase; NEMO, NF-κB essential modifier; NNK, nicotine-derived 4-(methylnitrosamino)-1-(3-pyridyl)-1-butanone; UV, ultra violet; cIAP, cellular inhibitor of apoptosis; TNF, tumor necrosis factor; TRAF, TNF receptor-associated factor; SOD, superoxide dismutase; CAM, cell adhesion molecules; COX, cyclooxygenase; iNOS, inducible nitric oxide synthase; uPA, urokinase-type plasminogen activator; MMP, matrix metalloproteinase; VEGF, vascular endothelial cell growth factor; IL, interleukin; EGFR, epidermal growth factor receptor; CAPE, caffeic acid phenethyl ester; PBIT; S,S'-1,4-phenylene-bis(1,2-ethanediyl)bis-isothiourea; PDTC, pyrrolidine dithocar-

members bind to DNA, only p65, c-Rel, and RelB contain a transactivation domain. In most resting cells, NF- κ B is retained in the cytoplasm by binding to the inhibitory I κ B proteins (I κ B α , I κ B β , I κ B ϵ , p105, and p100), which blocks the nuclear localization sequences of NF- κ B [8]. NF- κ B is activated in response to a wide variety of stimuli that promote the dissociation of the I κ B α through phosphorylation, ubiquitination, and degradation, thus unmasking of the nuclear localization sequence of NF- κ B thereby allowing NF- κ B to enter the nucleus and bind κ B-regulatory elements [9]. The phosphorylation of I κ B α is a critical step in the pathway leading to NF- κ B activation, and this step is catalyzed by an I κ B α kinase (IKK) complex (molecular mass of 700 kDa) consisting of IKK- α , IKK- β , IKK- γ (also called NEMO), and other proteins yet to be identified [10].

Because of the critical role of NF- κB in cell survival, cell adhesion, inflammation, differentiation, and cell growth, it has been implicated in carcinogenesis. Cancer is a hyperproliferative disorder that results from tumor initiation and tumor promotion, which ultimately produces tumor metastasis. Several genes that are involved in cellular transformation, proliferation, invasion, and angiogenesis are regulated by NF- κB [11,12]. The focus of this review is to present the evidence that NF- κB can lead to tumorigenesis and, therefore, establishes that agents that suppress NF- κB can abrogate carcinogenesis. We also present evidence that suppression of NF- κB can sensitize tumor cells to chemotherapeutic agents.

1.1. NF- κB is activated by carcinogens and tumor promoters

Several recent studies indicate that NF- κ B is activated by various carcinogens and tumor promoters. These include benzo[a]pyrene, NNK, UV radiation, and phorbol esters [9,13–16]. For instance UV radiation has been shown to cause sunburn reactions (swelling, leukocyte infiltration, epidermal hyperplasia, and accumulation of proinflammatory cytokines) leading to skin cancer, and suppression of NF- κ B blocks the sunburn-induced damage [17].

1.2. NF- κB is needed for cell survival and cell proliferation

In 1996, four independent reports showed that activation of NF-κB promotes cell survival and downregulation of NF-κB sensitizes the cells to apoptosis induced by cytokines and chemotherapeutic agents [18–21]. How NF-κB activation promotes cell survival is becoming increasingly clear. The expression of several genes including bcl-2, $bcl-x_L$, cIAP, xIAP, TRAF1, TRAF2, SOD, and A2O, have been reported to be regulated by NF-κB and to mediate cell survival (Table 1). How these proteins enhance cell survival is not fully understood, but their role in blocking the apoptosis pathway has been demonstrated [3]. Studies

Table 1 Antiapoptotic genes products regulated by NF- κB^a

Bcl-2
$Bcl-x_L$
TNF receptor-associated factor-1 (TRAF1)
TNF receptor-associated factor-2 (TRAF2)
Mn superoxide dismutase (MnSOD)
Survivin
Cyclin D1
γ-Glutamylcysteine synthetase (γ-GCS)
Cellular inhibitor of apoptosis (cIAP-1)
cIAP-2
xIAP
A20
Cyclooxygenase-2
c-Myc

^a For references see [3].

from our laboratory and others have shown that expression of activated NF- κ B promotes cell proliferation and suppression of NF- κ B leads to abrogation of proliferation [22–24].

1.3. NF- κB regulates the genes involved in tumor cell invasion and angiogenesis

Invasion and angiogenesis are critical events for tumor metastasis. Various genes that are involved in tumor cell invasion and angiogenesis have also been found to be regulated by NF-κB. These include the cell adhesion molecules (ICAM-1, VCAM-1, ELAM-1), COX2, iNOS, uPA, MMP-9, MMP-2, VEGF, chemokines, and inflammatory cytokines (Table 2). Thus, the suppression of NF-κB activation will likely abrogate the expression of these genes and, thus, prevent tumor metastasis.

1.4. NF-κB is constitutively expressed in tumor cells

While it is clear from these description that NF- κB is needed for tumor cell proliferation, invasion, and angio-

Table 2 NF-κB-regulated genes involved in carcinogenesis

Gene product	References	
c-Myc	[34,35]	
Epidermal growth factor receptor (EGFR)	[36]	
Cell adhesion molecules	[37]	
Inducible nitric oxide synthase (iNOS)	[38-40]	
Vascular endothelial cell growth factor (VEGF)	[39,41]	
Interleukin-6	[42]	
Cyclin D1	[43,44]	
Matrix metalloproteinase-9 (MMP-9)	[45]	
Cyclooxygenase-2 (COX2)	[39,46,47]	
Interleukin-8	[48]	
Urokinase-type plasminogen activator (uPA)	[49]	
Bcl-2	[50,51]	
$Bcl-x_L$	[50,52]	
Matrix metalloproteinase-2 (MMP-2)	[45]	
Gro 1	[53]	

genesis, tumor cells have been found to constitutively express the activated form of NF-κB. Several different tumor cell types, including leukemia, lymphoma, myeloma, melanoma, prostate, colon, breast, pancreas, and head and neck squamous cell carcinoma cell lines have been reported to express constitutively active NF-kB [23,25–27]. We and others have shown that samples obtained from cancer patients also exhibit constitutive NF-κB [24]. What causes the constitutive activation of NF-κB is not fully understood, but the roles of TNF, IL-1, pH, and hypoxia has been demonstrated. Our laboratory has shown that constitutive NF-kB in T cell cutaneous lymphoma [28] and in acute myelogenous leukemia [29] is due to constitutive expression of TNF and IL-1, respectively. Suppression of TNF and IL-1 production was found to downregulate the expression of active NF-κB, which

correlated with inhibition of proliferation of these tumor cells.

1.5. NF-kB activation is suppressed by chemopreventive agent

How chemopreventive agents suppress tumorigenesis is not fully understood. Several assays have been developed to determine the chemopreventive ability of an agent [30]. We have noted that most agents that have chemopreventive effects suppress the activation of NF- κ B. These include curcumin, resveratrol, emodin, green tea polyphenols, silymarin, β -lapachone, caffeic acid phenethyl ester, and sulindac (Table 3). Since NF- κ B regulates the expression of numerous genes that are involved in carcingenesis (Table 2), the suppression of expression of these genes

Table 3 Chemopreventive agents that block NF- κB activation

Chemopreventive agent	Source	Chemical name	References
Curcumin	Curcuma longa	(<i>E,E</i>)-1,7-Bis(4-hydroxy-3-methoxy-phenyl)-1,6-heptadiene-3,5-dione; diferuloylmethane	[23,32,40,54]
Retinoids	Vitamin A	N-(4-Hydroxyphenyl) retinamide; all- <i>trans</i> retinoic acid	[55]
Capsaicin	Capsicum sp.	<i>N</i> -(4-Hydroxy-3-methoxybenzyl)-8-methylnon- <i>trans</i> -6-enamide	[54,56]
Vitamin E	Plants	α-Tocopherol; 5,7,8-trimethyltocol	[57]
Quercetin	Rhododendron cinnabarinum, widely distributed in plant kingdom in rinds and barks	3,3',4',5,7-Pentahydroxyflavone	[58]
Dihydroxy vitamin D3	Fish liver oils	1α,25-Dihydroxycholecalciferol	[57]
Resveratrol	Phytoalexin found in a variety of plants, abundant in <i>Polygonum</i> cuspidatum and grape fruits	trans-3,5,4'-Trihydroxystilbene	[33,59]
Silymarin	Silybum marianum	Silybin	[60]
Lapachone	Heartwood of bignoniaceous plants, e.g. Lapacha tree	β-Lapachone	[61]
Sulindac	Synthetic	(Z)-Fluoro-2-methyl-1- [[4-(methyl-sulfinyl)phenyl]methylene]- 1H-indene-3-acetic acid	[62]
Celecoxib	Synthetic	1,2-Diarylpyrrole	[47]
Tea polyphenols	Green or black tea	(-)-Epigallocatechin-3-gallate; theaflavin	[63]
Sulforaphane	Cruciferous vegetables, e.g. Broccoli	1-Isothiocyanato-(4R)-(methylsulfinyl)butane	[46]
Aspirin	Synthetic	2-(Acetyloxy)benzoic acid	[64]
Caffeic acid phenethyl ester (CAPE)	Propolis from honeybee hives	3,4-Dihydroxycinnamic acid phenylethyl ester	[65]
S,S'-1,4-Phenylene-bis(1,2- ethanediyl)bis-isothiourea (PBIT)	Synthetic	<i>S,S'</i> -1,4-Phenylene-bis(1,2-ethanediyl)bisisothiourea (PBIT)	[66]
Pyrrolidine dithocarbamate (PDTC)	Synthetic	Pyrrolidine dithocarbamate	[67]
Anethole	Pimpinella anisum, camphor, fennel	<i>p</i> -Propenylanisole	[68]
Oleandrin	Nerium oleander	(3β,5β,16β)-16-(Acetyloxy)-3-[(2,6-di-deoxy-3- <i>O</i> -methyl-α-arabino-hexopyranosyl)oxy]-14- hydroxycard-20(22)-enolide	[69]
Wortmannin	Antibiotic from Penicillium wormanni	[IS-(1α,6bα,9aβ,11α,11bβ)]-11-(Acetyloxy)- 1,6b,7,8,9a,10,11,11b-octahydro-1- (methoxymethyl)-9a, 11b-dimethyl-3H-furo[4,3,2- de]indeno[4,5-h]-2-benzopyran-3,6,9-trione	[70]
Emodin	Aloe barbandensis, Polygonum cuspidatum	1,3,8-Trihydroxy-6-methyl-9,10-anthracenedione	[71]

Rationale for a combination therapy of cancer

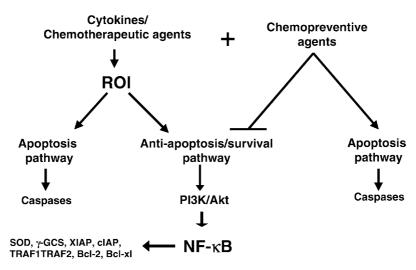


Fig. 1. Chemopreventive agents potentiate the effect of chemotherapeutic agents through downregulation of cell survival mechanisms. PI3K is phosphotidylinositol-3-kinase, SOD is superoxide dismutase, γ -GCS is gamma glutamylcysteine synthetase, and cIAP is cellular inhibitor of apoptosis.

through inhibition of NF- κB activation may be one of the mechanisms by which chemopreventive agents mediate their effects.

1.6. NF- κB is activated by chemotherapeutic agents leading to chemoresistance

It has been found that most chemotherapeutic agents activate NF- κ B. These include taxol, doxorubicin, daunorubicin, etoposide, vincristin, vinblastin, ara-C, anthralin, AZT, ciprofirate, cisplatin, haloperidol, methamphetamine, phenobarbital, temoxifen, and camptothecin [9]. Even gamma irradiation, commonly used to treat cancer patients, has also been found to activate NF- κ B [31]. The activation of NF- κ B can lead to resistance to apoptosis ordinarily induced by chemotherapy or radiation therapy. Thus, while activating apoptosis, the same agent can also activate NF- κ B, which can lead to antiapoptosis.

1.7. NF- κB suppression sensitizes the tumor cells to chemotherapeutic agents and to ionizing radiation

Most chemotherapeutic agents and ionizing radiation induce apoptosis through activation of various caspases. They also activate antiapoptosis through activation of NF- κ B, which leads to eventual resistance of tumor cells to therapy. Since most chemotherapeutic agents are known to suppress the activation of NF- κ B and NF- κ B-regulated gene expression, we propose the use of chemotherapeutic agents or gamma radiation in combination with chemopreventive agents for the therapy of cancer (Fig. 1). Besides blocking NF- κ B activation, chemopreventive agents such as curcumin and resveratrol are also known to induce

apoptosis [23,32,33]. Thus, unlike chemotherapeutic agents, chemopreventive agents induce apoptosis without activating the antiapoptosis pathway. Because most chemopreventive agents are natural plant-derived products, there is minimum toxicity associated with them. This provides an additional rationale for combination therapy.

2. Conclusions

Evidence presented above suggests that activation of NF-κB can lead to tumor cell proliferation, invasion, angiogenesis, and metastasis. Thus, suppression of NFκB in cancer cells may provide an additional target for prevention of cancer. NF-kB blockers can also be considered for the therapy of cancer, perhaps in combination with chemotherapeutic agents or gamma irradiation. Cancer is a multifactorial disease, and its treatment may also require multimodal therapy. In most instances when the diseases is diagnosed, numerous changes in the tumor have already taken place, thus creating a challenge for the physician to treat the patient. Most recent treatments for cancer have been a combination therapy consisting of chemotherapeutic agents, inhibitors of signal transduction and antibodies against various cell surface antigens. Therefore, it is reasonable to combine chemotherapeutic agents with chemopreventive agents for the treatment of cancer (Fig. 1).

Acknowledgments

This research was supported by grants from the Clayton Foundation, National Institute of Health (1P01 CA91844-1),

and by the Department of Defense (BC010610) to BBA. We will like to thank Walter Pagel for a careful review of the manuscript.

References

- Sen R, Baltimore D. Inducibility of kappa immunoglobulin enhancerbinding protein Nf-kappa B by a posttranslational mechanism. Cell 1986;47(6):921–8.
- [2] Aggarwal BB, Vilcek J. Tumor necrosis factor: structure, function and mechanism of action. New York: Marcel Dekker Inc, 1992.
- [3] Shishodia S, Aggarwal BB. Nuclear factor-kappa B activation: a question of life and death. J Biochem Mol Biol 2002;35:28–40.
- [4] Collins T, Read MA, Neish AS, Whitley MZ, Thanos D, Maniatis T. Transcriptional regulation of endothelial cell adhesion molecules: NF-kappa B and cytokine-inducible enhancers. FASEB J 1995;9(10): 800,000
- [5] Chen F, Castranova V, Shi X. New insights into the role of nuclear factor-kappaB in cell growth regulation. Am J Pathol 2001;159(2): 387–97.
- [6] Thanos D, Maniatis T. NF-kappa B: a lesson in family values. Cell 1995;80(4):529–32.
- [7] Baeuerle PA, Baltimore D. NF-kappa B: ten years after. Cell 1996; 87(1):13–20.
- [8] Baldwin Jr AS. The NF-kappa B and I kappa B proteins: new discoveries and insights. Annu Rev Immunol 1996;14:649–83.
- [9] Pahl HL. Activators and target genes of Rel/NF-kappaB transcription factors. Oncogene 1999;18(49):6853–66.
- [10] Karin M. The beginning of the end: IkappaB kinase (IKK) and NF-kappaB activation. J Biol Chem 1999;274(39):27339–42.
- [11] Kabrun N, Enrietto PJ. The Rel family of proteins in oncogenesis and differentiation. Semin Cancer Biol 1994;5(2):103–12.
- [12] Baldwin AS. Control of oncogenesis and cancer therapy resistance by the transcription factor NF-kappaB. J Clin Invest 2001;107(3):241-6.
- [13] Yan Z, Subbaramaiah K, Camilli T, Zhang F, Tanabe T, McCaffrey TA, Dannenberg AJ, Weksler BB. Benzo[a]pyrene induces the transcription of cyclooxygenase-2 in vascular smooth muscle cells. Evidence for the involvement of extracellular signal-regulated kinase and NFkappaB. J Biol Chem 2000;275(7):4949–55.
- [14] Rioux N, Castonguay A. The induction of cyclooxygenase-1 by a tobacco carcinogen in U937 human macrophages is correlated to the activation of NF-kappaB. Carcinogenesis 2000;21(9):1745–51.
- [15] Li N, Karin M. Ionizing radiation and short wavelength UV activate NF-kappaB through two distinct mechanisms. Proc Natl Acad Sci USA 1998;95(22):13012–7.
- [16] Baeuerle PA, Lenardo M, Pierce JW, Baltimore D. Phorbol-ester-induced activation of the NF-kappa B transcription factor involves dissociation of an apparently cytoplasmic NF-kappa B/inhibitor complex. Cold Spring Harb Symp Quant Biol 1988;53(Pt 2):789–98.
- [17] Abeyama K, Eng W, Jester JV, Vink AA, Edelbaum D, Cockerell CJ, Bergstresser PR, Takashima A. A role for NF-kappaB-dependent gene transactivation in sunburn. J Clin Invest 2000;105(12):1751–9.
- [18] Wu M, Lee H, Bellas RE, Schauer SL, Arsura M, Katz D, FitzGerald MJ, Rothstein TL, Sherr DH, Sonenshein GE. Inhibition of NF-kappaB/Rel induces apoptosis of murine B cells. EMBO J 1996; 15(17):4682–90.
- [19] Beg AA, Baltimore D. An essential role for NF-kappaB in preventing TNF-alpha-induced cell death. Science 1996;274(5288):782–4.
- [20] Wang CY, Mayo MW, Baldwin Jr AS. TNF- and cancer therapyinduced apoptosis: potentiation by inhibition of NF-kappaB. Science 1996;274(5288):784–7.
- [21] Van Antwerp DJ, Martin SJ, Kafri T, Green DR, Verma IM. Suppression of TNF-alpha-induced apoptosis by NF-kappaB. Science 1996;274(5288):787–9.

- [22] Rath PC, Aggarwal BB. Antiproliferative effects of IFN-alpha correlate with the downregulation of nuclear factor-kappa B in human Burkitt lymphoma Daudi cells. J Interferon Cytokine Res 2001;21(7):523–8.
- [23] Mukhopadhyay A, Bueso-Ramos C, Chatterjee D, Pantazis P, Aggar-wal BB. Curcumin downregulates cell survival mechanisms in human prostate cancer cell lines. Oncogene 2001;20(52):7597–609.
- [24] Garg A, Aggarwal BB. Nuclear transcription factor as a target for cancer drug development. Leukemia 2002;16:1053–68.
- [25] Mori N, Fujii M, Ikeda S, Yamada Y, Tomonaga M, Ballard DW, Yamamoto N. Constitutive activation of NF-kappaB in primary adult T-cell leukemia cells. Blood 1999;93(7):2360–8.
- [26] Bargou RC, Emmerich F, Krappmann D, Bommert K, Mapara MY, Arnold W, Royer HD, Grinstein E, Greiner A, Scheidereit C, Dorken B. Constitutive nuclear factor-kappaB-RelA activation is required for proliferation and survival of Hodgkin's disease tumor cells. J Clin Invest 1997;100(12):2961–9.
- [27] Bours V, Dejardin E, Goujon-Letawe F, Merville MP, Castronovo V. The NF-kappa B transcription factor and cancer: high expression of NF-kappa B- and I kappa B-related proteins in tumor cell lines. Biochem Pharmacol 1994;47(1):145–9.
- [28] Giri DK, Aggarwal BB. Constitutive activation of NF-kappaB causes resistance to apoptosis in human cutaneous T cell lymphoma HuT-78 cells. Autocrine role of tumor necrosis factor and reactive oxygen intermediates. J Biol Chem 1998;273(22):14008–14.
- [29] Estrov Z, Manna SK, Harris D, Van Q, Estey EH, Kantarjian HM, Talpaz M, Aggarwal BB. Phenylarsine oxide blocks interleukin-1betainduced activation of the nuclear transcription factor NF-kappaB inhibits proliferation, and induces apoptosis of acute myelogenous leukemia cells. Blood 1999;94(8):2844–53.
- [30] Steele VE, Sharma S, Mehta R, Elmore E, Redpath L, Rudd C, Bagheri D, Sigman CC, Kelloff GJ. Use of in vitro assays to predict the efficacy of chemopreventive agents in whole animals. J Cell Biochem Suppl 1996;26:29–53.
- [31] Prasad AV, Mohan N, Chandrasekar B, Meltz ML. Activation of nuclear factor kappa B in human lymphoblastoid cells by low-dose ionizing radiation. Radiat Res 1994;138(3):367–72.
- [32] Han SS, Chung ST, Robertson DA, Ranjan D, Bondada S. Curcumin causes the growth arrest and apoptosis of B cell lymphoma by downregulation of egr-1, c-myc, bcl-XL, NF-kappa B, and p53. Clin Immunol 1999;93(2):152–61.
- [33] Clement MV, Hirpara JL, Chawdhury SH, Pervaiz S. Chemopreventive agent resveratrol, a natural product derived from grapes, triggers CD95 signaling-dependent apoptosis in human tumor cells. Blood 1998;92(3):996–1002.
- [34] Nasi S, Ciarapica R, Jucker R, Rosati J, Soucek L. Making decisions through Myc. FEBS Lett 2001;490(3):153–62.
- [35] Slawin K, Kadmon D, Park SH, Scardino PT, Anzano M, Sporn MB, Thompson TC. Dietary fenretinide, a synthetic retinoid, decreases the tumor incidence and the tumor mass of ras + myc-induced carcinomas in the mouse prostate reconstitution model system. Cancer Res 1993;53(19):4461-5.
- [36] Boiko IV, Mitchell MF, Hu W, Pandey DK, Mathevet P, Malpica A, Hittelman WN. Epidermal growth factor receptor expression in cervical intraepithelial neoplasia and its modulation during an alphadifluoromethylornithine chemoprevention trial. Clin Cancer Res 1998;4(6):1383–91.
- [37] Tozawa K, Sakurada S, Kohri K, Okamoto T. Effects of anti-nuclear factor kappa B reagents in blocking adhesion of human cancer cells to vascular endothelial cells. Cancer Res 1995;55(18):4162–7.
- [38] Ambs S, Merriam WG, Bennett WP, Felley-Bosco E, Ogunfusika MO, Oser SM, Klein S, Shields PG, Billiar TR, Harris CC. Frequent nitric oxide synthase-2 expression in human colon adenomas: implication for tumor angiogenesis and colon cancer progression. Cancer Res 1998;58(2):334–41.
- [39] Marrogi AJ, Travis WD, Welsh JA, Khan MA, Rahim H, Tazelaar H, Pairolero P, Trastek V, Jett J, Caporaso NE, Liotta LA, Harris CC.

- Nitric oxide synthase cyclooxygenase 2 and vascular endothelial growth factor in the angiogenesis of non-small cell lung carcinoma. Clin Cancer Res 2000;6(12):4739–44.
- [40] Chan MM, Huang HI, Fenton MR, Fong D. In vivo inhibition of nitric oxide synthase gene expression by curcumin, a cancer preventive natural product with anti-inflammatory properties. Biochem Pharmacol 1998;55(12):1955–62.
- [41] Joseph IB, Vukanovic J, Isaacs JT. Antiangiogenic treatment with linomide as chemoprevention for prostate, seminal vesicle, and breast carcinogenesis in rodents. Cancer Res 1996;56(15):3404–8.
- [42] Suganuma M, Okabe S, Kurusu M, Iida N, Ohshima S, Saeki Y, Kishimoto T, Fujiki H. Discrete roles of cytokines TNF-alpha, IL-1, IL-6 in tumor promotion and cell transformation. Int J Oncol 2002; 20(1):131–6.
- [43] Langenfeld J, Kiyokawa H, Sekula D, Boyle J, Dmitrovsky E. Post-translational regulation of cyclin D1 by retinoic acid: a chemoprevention mechanism. Proc Natl Acad Sci USA 1997;94(22):12070–4.
- [44] Weinstein IB, Begemann M, Zhou P, Han EK, Sgambato A, Doki Y, Arber N, Ciaparrone M, Yamamoto H. Disorders in cell circuitry associated with multistage carcinogenesis: exploitable targets for cancer prevention and therapy. Clin Cancer Res 1997;3(12 Pt 2):2696–702.
- [45] Philip S, Bulbule A, Kundu GC. Osteopontin stimulates tumor growth and activation of promatrix metalloproteinase-2 through nuclear factor-kappa B-mediated induction of membrane type 1 matrix metalloproteinase in murine melanoma cells. J Biol Chem 2001;276(48): 44926–35.
- [46] Heiss E, Herhaus C, Klimo K, Bartsch H, Gerhauser C. Nuclear factor kappa B is a molecular target for sulforaphane-mediated anti-inflammatory mechanisms. J Biol Chem 2001;276(34):32008–15.
- [47] Oshima M, Murai N, Kargman S, Arguello M, Luk P, Kwong E, Taketo MM, Evans JF. Chemoprevention of intestinal polyposis in the Apcdelta716 mouse by rofecoxib, a specific cyclooxygenase-2 inhibitor. Cancer Res 2001;61(4):1733–40.
- [48] Kunz M, Hartmann A, Flory E, Toksoy A, Koczan D, Thiesen HJ, Mukaida N, Neumann M, Rapp UR, Brocker EB, Gillitzer R. Anoxiainduced up-regulation of interleukin-8 in human malignant melanoma. A potential mechanism for high tumor aggressiveness. Am J Pathol 1999;155(3):753–63.
- [49] Wang W, Abbruzzese JL, Evans DB, Chiao PJ. Overexpression of urokinase-type plasminogen activator in pancreatic adenocarcinoma is regulated by constitutively activated RelA. Oncogene 1999;18(32): 4554-63
- [50] Pena JC, Thompson CB, Recant W, Vokes EE, Rudin CM. Bcl-xL and Bcl-2 expression in squamous cell carcinoma of the head and neck. Cancer 1999;85(1):164–70.
- [51] Diaz GD, Paraskeva C, Thomas MG, Binderup L, Hague A. Apoptosis is induced by the active metabolite of vitamin D3 and its analogue EB1089 in colorectal adenoma and carcinoma cells: possible implications for prevention and therapy. Cancer Res 2000;60(8):2304–12.
- [52] Kondo S, Shinomura Y, Kanayama S, Higashimoto Y, Miyagawa JI, Minami T, Kiyohara T, Zushi S, Kitamura S, Isozaki K, Matsuzawa Y. Over-expression of bcl-xL gene in human gastric adenomas and carcinomas. Int J Cancer 1996;68(6):727–30.
- [53] Loukinova E, Chen Z, Van Waes C, Dong G. Expression of proangiogenic chemokine Gro 1 in low and high metastatic variants of Pam murine squamous cell carcinoma is differentially regulated by IL-1alpha EGF and TGF-beta1 through NF-kappaB dependent and independent mechanisms. Int J Cancer 2001;94(5):637–44.
- [54] Surh YJ, Han SS, Keum YS, Seo HJ, Lee SS. Inhibitory effects of curcumin and capsaicin on phorbol ester-induced activation of eukaryotic transcription factors NF-kappaB and AP-1. Biofactors 2000;12(1-4):107-12.
- [55] Delia D, Aiello A, Formelli F, Fontanella E, Costa A, Miyashita T, Reed JC, Pierotti MA. Regulation of apoptosis induced by the retinoid

- N-(4-hydroxyphenyl) retinamide and effect of deregulated bcl-2. Blood 1995;85(2):359-67.
- [56] Singh S, Natarajan K, Aggarwal BB. Capsaicin (8-methyl-N-vanillyl-6-nonenamide) is a potent inhibitor of nuclear transcription factor-kappa B activation by diverse agents. J Immunol 1996;157(10): 4412–20
- [57] Sokoloski JA, Hodnick WF, Mayne ST, Cinquina C, Kim CS, Sartorelli AC. Induction of the differentiation of HL-60 promyelocytic leukemia cells by vitamin E and other antioxidants in combination with low levels of vitamin D3: possible relationship to NF-kappaB. Leukemia 1997;11(9):1546–53.
- [58] Natarajan K, Manna SK, Chaturvedi MM, Aggarwal BB. Protein tyrosine kinase inhibitors block tumor necrosis factor-induced activation of nuclear factor-kappaB, degradation of IkappaBalpha, nuclear translocation of p65 and subsequent gene expression. Arch Biochem Biophys 1998;352(1):59–70.
- [59] Manna SK, Mukhopadhyay A, Aggarwal BB. Resveratrol suppresses TNF-induced activation of nuclear transcription factors NF-kappa B activator protein-1 and apoptosis: potential role of reactive oxygen intermediates and lipid peroxidation. J Immunol 2000;164(12): 6509–19.
- [60] Manna SK, Mukhopadhyay A, Van NT, Aggarwal BB. Silymarin suppresses TNF-induced activation of NF-kappa B c-Jun N-terminal kinase and apoptosis. J Immunol 1999;163(12):6800–9.
- [61] Manna SK, Gad YP, Mukhopadhyay A, Aggarwal BB. Suppression of tumor necrosis factor-activated nuclear transcription factor-kappaB activator protein-1, c-Jun N-terminal kinase and apoptosis by betalapachone. Biochem Pharmacol 1999;57(7):763–74.
- [62] Yamamoto Y, Yin MJ, Lin KM, Gaynor RB. Sulindac inhibits activation of the NF-kappaB pathway. J Biol Chem 1999;274(38): 27307–14.
- [63] Nomura M, Ma W, Chen N, Bode AM, Dong Z. Inhibition of 12-O-tetradecanoylphorbol-13-acetate-induced NF-kappaB activation by tea polyphenols, (—)-epigallocatechin gallate, and theaflavins. Carcinogenesis 2000;21(10):1885–90.
- [64] Stark LA, Din FV, Zwacka RM, Dunlop MG. Aspirin-induced activation of the NF-kappaB signaling pathway: a novel mechanism for aspirin-mediated apoptosis in colon cancer cells. FASEB J 2001; 15(7):1273-5.
- [65] Natarajan K, Singh S, Burke Jr TR, Grunberger D, Aggarwal BB. Caffeic acid phenethyl ester is a potent and specific inhibitor of activation of nuclear transcription factor NF-kappa B. Proc Natl Acad Sci USA 1996;93(17):9090-5.
- [66] Rao CV, Kawamori T, Hamid R, Reddy BS. Chemoprevention of colonic aberrant crypt foci by an inducible nitric oxide synthaseselective inhibitor. Carcinogenesis 1999;20(4):641–4.
- [67] Li JJ, Westergaard C, Ghosh P, Colburn NH. Inhibitors of both nuclear factor-kappaB and activator protein-1 activation block the neoplastic transformation response. Cancer Res 1997;57(16):3569–76.
- [68] Chainy GB, Manna SK, Chaturvedi MM, Aggarwal BB. Anethole blocks both early and late cellular responses transduced by tumor necrosis factor: effect on NF-kappaB, AP-1, JNK, MAPKK and apoptosis. Oncogene 2000;19(25):2943–50.
- [69] Manna SK, Sah NK, Newman RA, Cisneros A, Aggarwal BB. Oleandrin suppresses activation of nuclear transcription factor-kappaB, activator protein-1, and c-Jun NH2-terminal kinase. Cancer Res 2000;60(14):3838–47.
- [70] Manna SK, Aggarwal BB. Wortmannin inhibits activation of nuclear transcription factors NF-kappaB and activated protein-1 induced by lipopolysaccharide and phorbol ester. FEBS Lett 2000;473(1): 113–8.
- [71] Kumar A, Dhawan S, Aggarwal BB. Emodin (3-methyl-1,6,8-trihydroxyanthraquinone) inhibits TNF-induced NF-kappaB activation, IkappaB degradation, and expression of cell surface adhesion proteins in human vascular endothelial cells. Oncogene 1998;17(7):913–8.